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Synthesis of Galactopyranosyl Substituted Porphyrins

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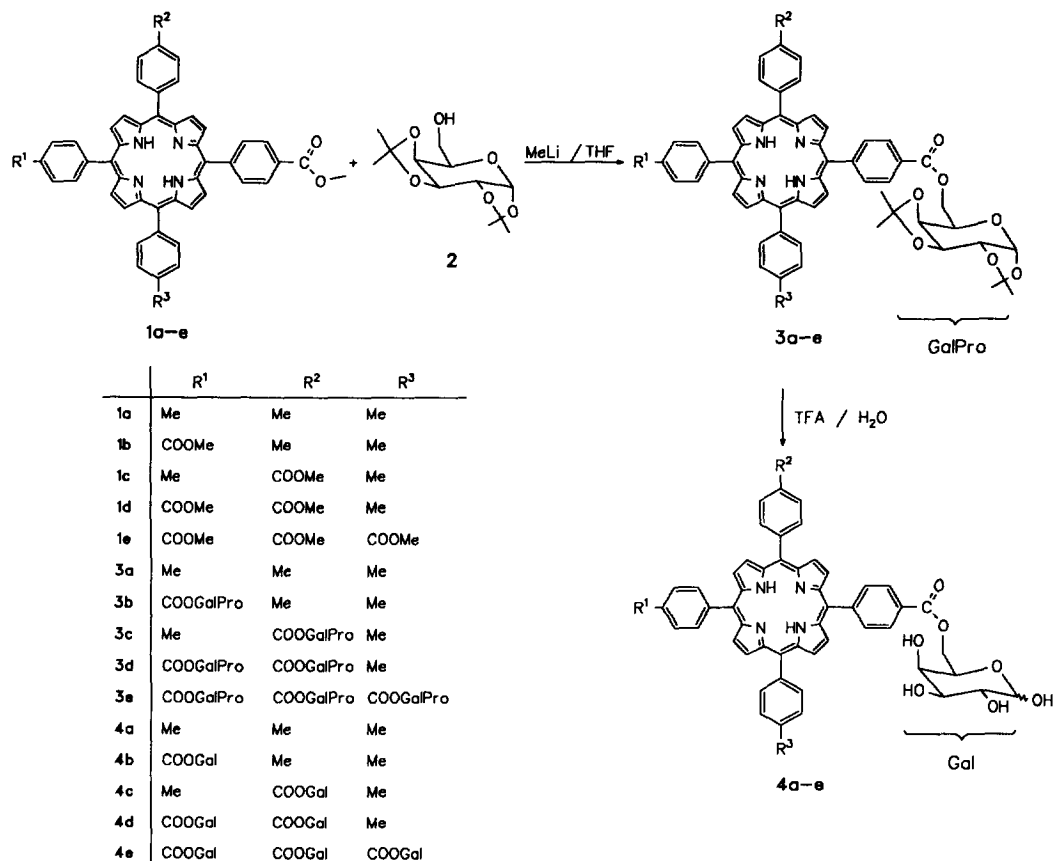
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Abstract: The synthesis of several galactopyranosyl substituted porphyrins is described. Starting from porphyrin carboxylic esters (1) di-O-isopropylidene- α -D-galactopyranosyl substituted porphyrins 3a - 3e were synthesised by a transesterification procedure. Treatment of 3a - 3e with TFA/ water leads to the formation of galactopyranosyl substituted porphyrins 4a - 4e in excellent yield. Furthermore porphyrin 3e can be synthesised by reaction of di-O-isopropylidene- α -D-galactopyranosyl substituted benzaldehyde 7 with pyrrole. Aldehyde 7 was synthesised by a CsF catalysed condensation reaction of 6-deoxy-6-bromo-di-O-isopropylidene- α -D-galactopyranose 6 with 4-formyl benzoic acid.

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One of the most interesting applications of porphyrins is photodynamic therapy (PDT) of tumours.¹ This treatment based on the interaction of the porphyrinic sensitizer with visible light, thus producing singlet oxygen directly in the cancer cell that causes cell death.² Today only a purified form of hematoporphyrin derivative (HpD), Photofrin II is used as photosensitizer in clinical studies and therapy. The main disadvantage of Photofrin II is its unknown composition of different porphyrins.³ Therefore the synthesis of well-defined porphyrins is of great interest for the development of new photosensitizers for use in PDT.⁴ One of the requirements a new photosensitizer has to fulfill is solubility in water. Therefore the synthesis of carbohydrate-substituted porphyrins is very interesting regarding PDT. Furthermore, a specific affinity of several carbohydrate substituted porphyrins to cancer cells was reported.⁵ Therefore, a number of carbohydrate-substituted porphyrins were synthesised.⁶ We now report here a new synthetic procedure for the synthesis of porphyrins bearing 1-4 carbohydrate substituents.

We recently reported the synthesis of porphyrins bearing long alkyl chains and cholesterol subunits by a transesterification procedure.⁷ Herein we describe a modified procedure of this methodology for the synthesis of carbohydrate substituted porphyrins. In this modified procedure porphyrinic carboxylic esters 1 were treated with 1,2,3,4-di-O-isopropylidene- α -D-galactopyranose (2) and methyl lithium in THF (Scheme 1). The reaction proceeds in good yield (28 - 50%) at 80 - 90 °C for the synthesis of compounds 3a - 3c. Synthesis of 3d and 3e, respectively, was performed in lower yield (7%) due to the increased formation of

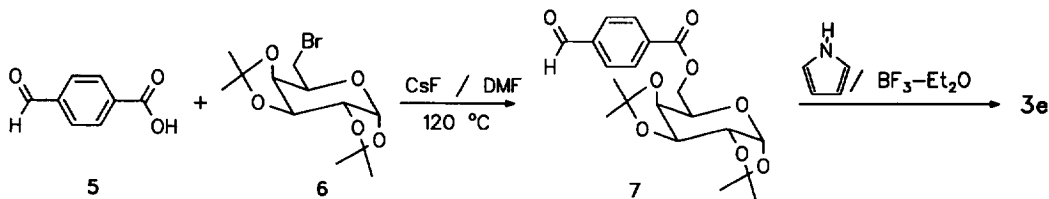


Scheme 1

porphyrin carboxylic acids. Formation of these side-products can be explained by direct saponification of the starting compounds **1a - 1e** or by thermal decomposition of products **3a - 3e**. Due to the extreme water free reaction conditions a direct saponification of starting compounds is unlikely. However, investigation of the thermostability of compounds **3a - 3e** showed, that these compounds are unstable and decompose to the observed side-products. We therefore carried out the reaction at lower temperature (40 °C). No formation of carboxylic acids was obtained under these reaction conditions; but unfortunately compounds **3a - 3e** were obtained in lower yield at this temperature.

Alternatively, compound **3e** can be easily synthesised in nearly 20% yield by reacting aldehyde **7** with pyrrole. The di-O-isopropylidene- α -D-galactopyranosyl substituted aldehyde **7** was synthesised from **6** and 4-formyl benzoic acid (Scheme 2) in a CsF catalysed condensation reaction.⁸ We also treated aldehyde **7** in a mixed condensation reaction with *p*-tolyl aldehyde and pyrrole using Lindsey's procedure.⁹

Although all porphyrins **3a** - **3e** were formed in this reaction the yields were very low (**3a** was formed in only 8% yield) compared with the yields obtained by the transesterification procedure.



Scheme 2

Deprotection of compounds **3a** - **3e** was performed in excellent yield (92 - 98%) with TFA/ H₂O (9:1) at room temperature. After neutralisation of the reaction mixture with Na₂CO₃ solution the porphyrins precipitated and were isolated by filtration. Purification of the deprotected porphyrins **4a** - **4d** was performed by column chromatography on silica gel using CH₂Cl₂/ MeOH (9:1) as eluent. TLC and NMR-spectroscopic studies showed, that all porphyrins **4a** - **4e** were formed as mixtures of α - and β -isomers.¹⁰

Identification of the synthesised carbohydrate substituted porphyrin derivatives was performed by ¹H NMR and ¹³C NMR spectroscopy and FAB spectroscopy. The obtained analytical data are also in accordance with the proposed structures.

The synthesised porphyrins exhibit very different solubility properties. Compound **4a** is soluble in CH₂Cl₂ or CHCl₃, whereas compounds **4b** and **4c**, respectively, show better solubility in a mixture of CHCl₃ and methanol. In contrast to this **4c** is only soluble in methanol. Compound **4e** is only soluble in DMSO or in water or methanol without aggregation up to concentrations of approximately 10⁻⁴ M, as indicated by absorption spectroscopy.

EXPERIMENTAL SECTION

NMR spectra were obtained in CDCl₃ or DMSO-d₆ and recorded with a Varian XL 200 spectrometer or a Bruker DMX-500 spectrometer. Chemical shift values were given in ppm relative to TMS. Mass spectra were measured with a VG-Analytical VG70:250 E instrument. Electronic spectra were recorded on a Kontron Uvikon 860 instrument. IR spectra were recorded on a Shimadzu IR-435 spectrometer. Melting points were measured on a Büchi 510 apparatus and are uncorrected. Column chromatography was carried out with Merck silica gel mesh size 0.060 - 0.2 mm.

Synthesis of 6'-deoxy-1',2':3',4'-di-O-isopropylidene- α -D-galactopyranosyl-4-formyl-benzoate (7)

To a solution of 1.30 g (4.02 mmol) **6** in 4 ml DMF 2.00 g (13.2 mmol) CsF and 336 mg (2.24 mmol) 4-formyl benzoic acid were added. The reaction mixture was stirred for 2 days at 120 °C. Then 40 ml CH₂Cl₂ were added. The mixture was poured into saturated NaHCO₃ solution (50 ml). The organic layer was separated and washed with water and dried (Na₂SO₄). The solvent was evaporated and the residue chromatographed on a silica gel column using CH₂Cl₂/methanol (98:2) as eluent. Yield: 616 mg (70%) **7** (colourless oil).- ¹H NMR (CDCl₃): δ = 1.35, 1.37, 1.70, 1.73 (4 s, 12 H, C(CH₃)₂), 4.10 - 4.81 (m, 6 H, 2', 3', 4', 5', 6'-H), 5.60 (d, J = 5.4 Hz, 1 H, 1'-H), 7.95 (d, J = 8.3 Hz, 2 H, H_o), 8.24 (d, J = 8.3 Hz, 2 H, H_m), 10.12 (s, 1 H, CHO).- MS (70 eV): m/e = 394 (M⁺ + 2).- Anal. Calc. for C₂₀H₂₄O₈ (392.40): C 61.22 H 6.16. Found: C 60.76 H 6.61.

5-(p-1',2':3',4'-Di-O-isopropylidene- α -D-galactopyranoloxycarbonylphenyl)-10,15,20-tris(p-tolyl)-porphyrin (3a)

In a dry round bottom flask 1.27 g (4.88 mmol) 1,2,3,4-di-O-isopropylidene- α -D-galactopyranose (**2**) were added under nitrogen to 10 ml of dry THF. The mixture was cooled down to 1-2 °C. Then 3.00 ml (4.80 mmol) of a 1.6 M solution of methyllithium in ether were added. The reaction mixture was warmed up to room temperature and 241 mg (0.34 mmol) of porphyrin **1a** were added. The reaction mixture was stirred for 8 h at 80 - 90 °C. Then 70 ml CH₂Cl₂ were added and the reaction mixture was poured into water (40 ml). The organic layer was isolated and dried (Na₂SO₄). The solvent was evaporated and the residue chromatographed on a silica gel column using CH₂Cl₂/methanol (98:2) as eluent. Yield: 114 mg (38%) **3a**. ¹H NMR (CDCl₃): δ = -2.78 (br s, 2 H, NH), 1.38, 1.42, 1.55, 1.63 (4 s, 12 H, C(CH₃)₂), 2.70 (s, 9 H, CH₃), 4.36 - 4.49, 4.64 - 4.76 (2 m, 6 H, 2', 3', 4', 5', 6'-H), 5.66 (d, J = 5.0 Hz, 1 H, 1'-H), 7.55 (d, J = 7.9 Hz, 6 H, H_m), 8.09 (d, J = 7.9 Hz, 6 H, H_o), 8.30 (d, J = 8.3 Hz, 2 H, H_o), 8.78 (d, 8.3 Hz, 2 H, H_m), 8.79 (d, J = 4.9 Hz, 2 H, 3-, 7-H), 8.87 (s, 4 H, 12-, 13-, 17-, 18-H), 8.88 (d, 2 H, 8-H).- ¹³C NMR (CDCl₃): δ = 21.55 (q, CH₃), 24.61, 25.05, 26.10, 26.16 (4 q, C(CH₃)₂), 64.24 (t, C-6'), 66.26 (d, C-5'), 70.58, 70.81 (2 d, C-3', C-4'), 71.25 (d, C-2'), 96.46 (d, C-1'), 108.92, 109.80 (2 s, C(CH₃)₂), 118.29, 120.41, 120.69 (3 s, C_{meso}), 127.44, 128.04 (2 d, C_m, C_m'), 129.40 (s, C_p'), 131.26 (br d, C _{β}), 134.51, 134.61 (2 d, C_o, C_o'), 137.41 (s, C_p), 139.15 (s, C_i), 147.29 (s, C_i'), 166.73 (s, CO).- IR (KBr): ν = 1717 cm⁻¹ (CO).- UV-Vis. (CH₂Cl₂): λ = 419, 515, 551, 591, 646 nm.- FAB-MS: m/e = 944 (M⁺ + 1), 701, 656.- Anal. Calc. for C₆₀H₅₄N₄O₇ (943.1): C 76.41, H 5.77, N 5.94. Found: C 76.43, H 5.78, N 5.59.

5-15-Bis(p-1',2':3',4'-di-O-isopropylidene- α -D-galactopyranoloxycarbonylphenyl)-10,20-bis(p-tolyl)-porphyrin (3b)

According to the procedure given above, 44 mg (0.058 mmol) porphyrin **1b**, 625 mg (2.40 mmol) **2** and 1.00 ml (1.60 mmol) methyllithium were treated. Reaction time 2.5 h. Yield: 35 mg (50%) **3b**.- ¹H NMR (CDCl₃): δ = -2.79 (br s, 2 H, NH), 1.38, 1.42, 1.56, 1.63 (4 s, 24 H, C(CH₃)₂), 2.70 (s, 6 H, CH₃), 4.36 - 4.49, 4.64 - 4.76 (2 m, 12 H, 2', 3', 4', 5', 6'-H), 5.66 (d, J = 4.9 Hz, 2 H, 1'-H), 7.56 (d, J = 7.8 Hz, 4 H, H_m), 8.09 (d, J = 7.8 Hz, 4 H, H_o), 8.30 (d, J = 8.3 Hz, 4 H, H_o), 8.45 (d, J = 8.3 Hz, 4 H, H_m), 8.79 (d, J = 4.8 Hz, 4 H, 3-, 7-, 13-, 17-H), 8.90 (d, J = 4.9 Hz, 4 H, 2-, 8-, 12-, 18-H).- ¹³C NMR (CDCl₃): δ = 21.55 (q, CH₃), 24.58, 25.02, 26.06, 26.26 (4 q, C(CH₃)₂), 64.24 (t, C-6'), 66.23 (d, C-5'), 70.55, 70.80 (2 d, C-3', C-4'), 71.25 (d, C-2'), 96.43 (d, C-1'), 108.92, 109.80 (2 s, C(CH₃)₂), 118.80, 120.69 (2 s, C_{meso}), 127.51, 128.01 (2 d, C_m, C_m'), 129.43 (C_p'), 131.20 (br d, C _{β}), 134.48, 134.52, (2 d, C_o, C_o'), 137.54 (s, C_p), 138.93 (s, C_i), 147.10 (s, C_i'), 166.66 (s, CO).- IR (KBr): ν = 1717 cm⁻¹ (CO).- UV-Vis. (CH₂Cl₂): λ = 419, 515, 551, 591, 646 nm.- FAB-MS: m/e = 1215 (M⁺)- Anal. Calc. for C₇₂H₇₀N₄O₁₄ (1215.4): C 71.15 H 5.81 N 4.61. Found: C 71.12 H 5.87 N 4.54.

5,10-Bis(p-1',2':3',4'-di-O-isopropylidene- α -D-galactopyranoloxycarbonylphenyl)-15,20-bis(p-tolyl)-porphyrin (3c)

According to the procedure given above 44 mg (0.058 mmol) porphyrin 1c, 250 mg (0.96 mmol) 2 and 0.57 ml (0.91 mmol) methyllithium were treated. Reaction time 2.5 h. Purification of compound 3c was performed by column chromatography on silica gel using CH₂Cl₂/ methanol (97:3) as eluent. Yield: 20 mg (28%) 3c.- ¹H NMR (CDCl₃): δ = -2.78 (br s, 2 H, NH), 1.38, 1.42, 1.56, 1.63 (4 s, 24 H, C(CH₃)₂), 2.71 (s, 6 H, CH₃), 4.36 - 4.50, 4.64 - 4.76 (2 m, 12 H, 2', 3', 4', 5', 6'-H), 5.66 (d, 4.9 Hz, 2 H, 1'-H), 7.56 (d, *J* = 7.8 Hz, 4 H, H_m), 8.10 (d, *J* = 7.8 Hz, 4 H, H_o), 8.30 (d, *J* = 7.8 Hz, 4 H, H_o), 8.45 (d, *J* = 8.3 Hz, 4 H, H_m), 8.78 (d, *J* = 4.9 Hz, 2 H, 3-H, 12-H), 8.80 (s, 2 H, 7-H, 8-H), 8.88 (s, 2 H, 17-H, 18-H), 8.89 (d, *J* = 6.3 Hz, 2 H, 2-H, 13-H).- ¹³C NMR (CDCl₃): δ = 21.55 (q, CH₃), 24.61, 25.05, 26.06, 26.15 (4 q, C(CH₃)₂), 64.24 (t, C-6'), 66.25 (d, C-5'), 70.58, 70.83 (2 d, C-3', C-4'), 71.28 (d, C-2'), 96.45 (d, C-1'), 108.92, 109.83 (2 s, C(CH₃)₂), 118.04, 118.57, 120.97 (3 s, C_{meso}), 127.50, 128.07 (2 d, C_m, C_m), 129.46 (s, C_p), 131.13 (br d, C _{β}), 134.51, 134.57 (2 d, C_o, C_o), 137.54 (s, C_p), 139.02 (s, C_i), 147.00 (s, C_i), 166.69 (s, CO).- IR (KBr): ν = 1717 cm⁻¹ (CO).- UV-Vis (CH₂Cl₂): λ = 419, 515, 551, 591, 646 nm.- FAB-MS: *m/e* = 1214 (M⁺).- Anal. Calc. for C₇₂H₇₀N₄O₁₄ (1215.4): C 71.15 H 5.81 N 4.61. Found: C 71.13 H 5.74 N 4.70.

5,10,15-Tris(p-1',2':3',4'-di-O-isopropylidene- α -D-galactopyranoloxycarbonylphenyl)-20-(p-tolyl)-porphyrin (3d)

According to the procedure given above 240 mg (0.30 mmol) porphyrin 1d, 1.60g (6.14 mmol) 2 and 3.8 ml (6.08 mmol) methyllithium solution were treated. Reaction time 5 h. Purification of compound 3d was performed by column chromatography on silica gel using CH₂Cl₂/ methanol (97:3) as eluent. Yield. 31 mg (7%) 3d.- ¹H NMR (CDCl₃): δ = -2.79 (br s, 2 H, NH), 1.38, 1.42, 1.56, 1.63 (4 s, 36 H, C(CH₃)₂), 2.71 (s, 3 H, CH₃), 4.36 - 4.50, 4.62 - 4.76 (2 m, 18 H, 2', 3', 4', 5', 6'-H), 5.67 (d, *J* = 4.8 Hz, 3 H, 1'-H), 7.57 (d, *J* = 7.8 Hz, 2 H, H_m), 8.10 (d, *J* = 7.8 Hz, 2 H, H_o), 8.31 (d, *J* = 8.3 Hz, 6 H, H_o), 8.47 (d, *J* = 8.3 Hz, 6 H, H_m), 8.81 (d, *J* = 4.8 Hz, 2 H, 3-H, 17-H), 8.84 (s, 4 H, 7-H, 8-H, 12-H, 13-H), 8.92 (s, 2 H, 17-H, 18-H), 8.89 (d, *J* = 4.9 Hz, 2 H, 2-H, 18-H).- ¹³C NMR (CDCl₃): δ = 21.45 (q, CH₃), 24.58, 25.02, 26.06, 26.13 (4 q, C(CH₃)₂), 64.27 (t, C-6'), 66.23 (d, C-5'), 70.55, 70.80 (d, C-3', C-4'), 71.25 (d, C-2'), 96.43 (d, C-1'), 108.92, 109.80 (2 s, C(CH₃)₂), 118.86, 119.11, 121.15 (3 s, C_{meso}), 127.54, 128.10 (2 d, C_m, C_m), 129.52, 131.23 (br d, C _{β}), 134.54, 134.86 (2 d, C_o, C_o), 137.63 (s, C_p), 139.12 (s, C_i), 146.91 (s, C_i), 166.63 (s, CO).- IR (KBr): ν = 1717 cm⁻¹. UV-Vis (CH₂Cl₂): λ = 419, 515, 550, 590, 647 nm.- FAB-MS: *m/e* = 1488 (M⁺).- Anal. Calc. for C₈₄H₈₆N₄O₂₁ (1487.6): C 67.82 H 5.83 N 3.98. Found: C 67.50 H 6.27 N 3.75.

5,10,15,20-Tetrakis(p-1',2':3',4'-di-O-isopropylidene-D-galactopyranoloxycarbonylphenyl)-porphyrin (3e)

Method A:

According to the procedure given for 3a 150 mg (0.18 mmol) porphyrin 1e, 1.38 g (5.30 mmol) 2 and 3.3 ml (5.28 mmol) methyllithium solution were treated. Reaction time 4.5 h. Purification of 3e was performed on a silica gel column using CH₂Cl₂/ methanol (97:3) as eluent. Yield: 17 mg (5%) 3e.- ¹H NMR (CDCl₃): δ = -2.78 (br s, 2 H, NH), 1.39, 1.43, 1.56, 1.64 (4 s, 48 H, C(CH₃)₂), 4.37 - 4.50, 4.65 - 4.77 (2 m, 24 H, 2', 3', 4', 5', 6'-H), 5.67 (d, *J* = 5.4 Hz, 4 H, 1'-H), 8.31 (d, *J* = 6.9 Hz, 8 H, H_o), 8.48 (d, *J* = 8.4 Hz, 8 H, H_m), 8.85 (s, 8 H, H _{β}).- ¹³C NMR (CDCl₃): δ = 24.58, 25.02, 26.06, 26.13 (4 q, C(CH₃)₂), 64.27, (t, C-6'), 66.23 (d, C-5'), 70.55, 70.80 (2 d, C-3', C-4'), 71.25 (d, C-2'), 96.43 (d, C-1'), 108.92, 109.80 (2 s, C(CH₃)₂), 119.40 (s, C_{meso}), 128.14 (d, C_m), 129.62 (s, C_p), 131.25 (C _{β}), 134.54 (d, C_o), 146.71 (s, C_i), 166.60 (s, CO).- IR (KBr): ν = 1718 cm⁻¹. UV-Vis (CH₂Cl₂): λ = 419, 514, 549, 644.- FAB-MS: *m/e* = 1759 (M⁺).- Anal. Calc. for C₉₆H₁₀₂N₄O₂₈ (1759.9): C 65.53 H 5.81 N 3.28. Found: C 65.56 H 5.85 N 3.19.

Method B:

To 500 ml dry CH₂Cl₂ 1.77 g 7 (4.49 mmol) and 335 mg (0.50 mmol) pyrrole were added. Then 1 ml ethanol and 63 μ l BF₃·EtO₂ (50% solution) were added. The reaction mixture was stirred for 1 h at room temperature. Then 1.23 g (5 mmol) p-chloranile were added and the reaction mixture neutralised with NEt₃. The mixture was refluxed for 1 h. The solvent was evaporated and the residue chromatographed on a silica gel column using CH₂Cl₂/methanol (99.5:0.5). The obtained crude porphyrinic fraction was purified by a MPLC on silica gel using CH₂Cl₂/methanol (99.8:0.2) as eluent. Yield: 43.6 mg (19.8%) 3e.

Synthesis of 5-(p- α / β -D-galactopyran-6'-oloxycarbonylphenyl)-10,15,20-tris(p-tolyl)-porphyrin (4a)

Porphyrin 3a (28 mg, 0.032 mmol) was added to a mixture of CF₃COOH and water (9:1). The reaction mixture was stirred for 20 min at room temperature. Dichloromethane (20 ml) was added and the mixture cooled to 0°C. Then the mixture was poured into water (50 ml) and neutralised with Na₂CO₃ solution. The organic layer was separated and washed with water (30 ml) and dried (Na₂SO₄). The solvent was evaporated and the residue chromatographed on a silica gel column using CH₂Cl₂/methanol (9:1) as eluent. Yield 26 mg (94%) 4a. ¹H NMR (DMSO-d₆): δ = -2.92 (br s, 2 H, NH), 2.65 (s, 9 H, CH₃), 3.60 - 4.15 (m, 4 H), 4.29 - 5.00 (m, 4 H), 5.05 - 5.14 (m, 1 H, 1'-H_{ax}), 5.25 - 5.41 (m, 1 H), 6.37 (d, J = 4.9 Hz, 1 H, OH), 6.71 (d, J = 7.3 Hz, 1 H, OH), 7.59 (d, J = 7.8 Hz, 6 H, H_m), 8.07 (d, J = 7.8 Hz, 6 H, H_o), 8.31 - 8.48 (m, 4 H, H_o, H_m), 8.74 - 8.93 (m, 8 H, H_p). ¹³C NMR (DMSO-d₆): δ = 20.95 (q, CH₃), 64.75 (t, C-6'), 67.52 (d), 67.60 (d), 68.50 (d), 68.61 (d), 71.75 (d), 78.00 (d), 78.74 (d), 79.35 (d), 92.50 (d, α C-1'), 97.47 (d, β C-1'), 118.13 (s, C-5), 120.06 (s, C-10, C-20), 120.31 (s, C-15), 127.51, 127.69 (2 d, C_m, C_m), 129.27 (s, C_p), 131.32 (br d, C_p), 134.01, 134.42 (2d, C_o, C_o), 137.22 (s, C_p), 138.14 (s, C_i), 146.09 (s, C_i), 165.72 (s, CO).- IR (KBr): ν = 3395 (OH), 1700 (CO) cm⁻¹.- UV-Vis (CH₂Cl₂): λ = 418, 516, 550, 592, 647 nm.- FAB-MS: m/e = 863 (M⁺ + 1).- Anal. Calc. for C₅₄H₄₆N₄O₇ (863.0): C 75.16 H 5.37 N 6.49. Found: C 75.47 H 5.51 N 6.63.

5,15-Bis(p- α / β -galactopyran-6'-oloxycarbonylphenyl)-10,20-bis(p-tolyl)porphyrin (4b)

According to the procedure given for 4a, 35 mg (29 μ mol) porphyrin 3b were reacted. The reaction mixture was poured into water (50 ml) and neutralised with 20% Na₂CO₃ solution. The precipitated porphyrin 4b was isolated by filtration (0.45 μ m microfilter) washed with water and ethanol and dried. Yield: 28 mg (92%) 4b. ¹H NMR (DMSO-d₆): δ = -2.95 (br s, 2 H, NH), 2.65 (s, 6 H, CH₃), 3.60 - 4.10 (m, 8 H), 4.28 - 4.60 (m, 8 H), 4.80 - 4.88 (m, 2 H), 4.95 - 5.10 (m, 1 H, 1'-H_{ax}), 5.20 - 5.35 (m, 1 H), 6.35 (m, 1H, OH), 6.68 (m, 1 H, OH), 7.60 (d, J = 6.7 Hz, 4 H, H_m), 8.07 (d, J = 6.7 Hz, 4 H, H_o), 8.33 - 8.45 (m, 8 H, H_o-H_m), 8.80 (br s, 4 H, H_p), 8.88 (br s, 4 H, H_p). ¹³C NMR (DMSO): δ = 21.32 (q, CH₃), 65.02 (t, C-6'), 65.21 (t, C-6'), 67.88 (d), 68.14 (d), 68.98 (d), 69.01 (d), 69.50 (d), 69.61 (d), 69.99 (d), 72.27 (d), 72.31 (d), 73.46 (d), 81.77 (d), 83.04 (d), 93.11 (d, α C-1'), 97.88 (d, β C-1'), 102.19 (d), 119.05 (s, C-5, C-15), 120.71 (s, C-10, C-20), 127.95 (d, C_m), 128.00 (d, C_m), 129.68 (s, C_p), 133.00 (br d, C_p), 134.45 (d, C_o), 134.81 (d, C_o), 137.74 (s, C_p), 138.44 (s, C_i), 146.46 (s, C_i), 166.16 (s, CO).- IR (KBr): ν = 3400 (OH), 1705 (CO) cm⁻¹.- UV-Vis (CH₂Cl₂/methanol): λ = 420, 515, 553, 592, 649 nm.- Anal. Calc. for C₆₀H₅₄N₄O₁₄ (1055.1): C 68.30 H 5.16 N 5.31. Found: C 68.13 H 5.21 N 5.19.

5-10-Bis(p- α / β -D-galactopyran-6'-oloxycarbonylphenyl)-15,20-bis(p-tolyl)porphyrin (4c)

According to the procedure given for the synthesis of 4a, 48 mg (0.039 mmol) 3c were reacted. The reaction mixture was poured into water (60 ml) and neutralised with 20% Na₂CO₃ solution. The precipitated porphyrin 4c was isolated by filtration (0.45 μ m microfilter) washed with water and ethanol and dried. Yield 39 mg (95%) 4c. ¹H NMR (DMSO-d₆): δ = -2.92 (br s, 2 H, NH), 2.65 (s, 6 H, CH₃), 3.56 - 4.17 (m, 8 H), 4.25 - 4.67 (m, 6 H), 4.80 - 4.86 (m, 4 H), 4.95 - 5.11 (m, 2 H, 1'-H, H_{ax}), 5.20 - 5.35 (m, 1 H), 6.37 (d, J = 3.9 Hz, 2 H, OH), 6.71 (d, J = 6.9 Hz, 1 H, OH), 7.59 (br d, 4 H, H_m), 8.06 (br d, 4 H, H_o), 8.38 (br

8 H, H_{o,m}), 8.84 (br, 8 H, H_β).- ¹³C NMR (DMSO-d₆): δ = 20.95 (q, CH₃), 64.25 (t, CH₂), 66.00 (t, C-6'), 66.25 (t, C-6'), 67.40 (d), 67.62 (d), 68.45 (d), 68.90 (d), 69.05 (d), 71.65 (d), 72.74 (d), 75.77 (d), 81.25 (d), 82.63 (d), 92.65 (d, αC-1'), 97.47 (d, βC-1'), 118.39 (s, C-5, C-10), 120.56 (s, C-15, C-20), 127.54, 127.73 (2 d, C_m, C_{m'}), 129.18 (s, C_p), 131.48 (br d, C_β), 134.04, 134.42 (2 d, C_o, C_{o'}), 137.32 (s, C_p), 138.04 (s, C_i), 146.00 (s, C_i'), 165.65, 165.72 (2 s, CO).- IR (KBr): ν = 3395 (OH), 1705 cm⁻¹.- UV-Vis (CH₂Cl₂): λ = 418, 515, 550, 593, 645 nm.- FAB-MS: m/e = 1056 (M⁺ + 1).- Anal. Calc. for C₆₀H₅₄N₄O₁₄ (1055.1): C 68.30 H 5.16 N 5.31. Found: C 68.63 H 5.22 N 5.12.

5,10,15-Tris(p-α/β-D-galactopyran-6'-oloxycarbonylphenyl)-20-(p-tolyl)porphyrin (4d)

According to the procedure given for the synthesis of 4a, 23mg (0.016 mmol) 3d were reacted. The reaction mixture was poured into water (40 ml) and neutralised with 20% Na₂CO₃ solution. The precipitated porphyrin 4d was isolated by filtration (0.45 μm microfilter) washed with water and ethanol and dried. Yield: 19 mg (95%) 4d.- ¹H NMR (DMSO-d₆): δ = -2.96 (br s, 2 H, NH), 2.64 (s, 3 H, CH₃), 3.56 - 4.14 (m, 12 H), 4.22 - 4.67 (m, 10 H), 4.80 - 4.86 (m, 4 H), 4.95 - 5.11 (m, 2 H, 1'-H_o), 5.20 - 5.37 (m, 2 H), 6.35 (d, J = 5.3 Hz, 3 H, 1'-H_α), 6.70 (d, J = 6.8 Hz, 3 H, 1'-H_β), 7.61 (d, J = 7.8 Hz, 2 H, H_m), 8.07 (d, J = 7.8 Hz, 2 H, H_o), 8.37 (m, 12 H, H_{o'}, H_m), 8.85 (br, 8 H, H_β).- ¹³C NMR (DMSO-d₆): δ = 20.95 (q, CH₃), 64.65 (t, C-6'), 68.44 (d), 68.50 (d), 68.60 (d), 68.94 (d), 71.78 (d), 71.87 (d), 73.02 (d), 76.03 (d), 80.65 (d), 80.98 (d), 92.70 (d, αC-1'), 97.47 (d, βC-1'), 118.89, 120.05, 122.50 (3 s, C_{meso}), 127.60, 127.76 (2d, C_m, C_{m'}), 129.21 (s, C_p), 131.50 (br d, C_β), 134.07, 134.45 (2 d, C_o, C_{o'}), 137.25 (C_p), 139.95 (s, C_i), 145.90 (s, C_i'), 165.68, 165.72 (2 s, CO).- IR (KBr): ν = 3390 (OH), 1700 (CO) cm⁻¹.- UV-Vis (CH₂Cl₂/methanol): λ = 418, 517, 550, 595, 648 nm.- FAB-MS: m/e = 1247 (M⁺ + 1).- Anal. Calc. for C₆₆H₆₂N₄O₂₁ (1247.2): C 63.56 H 5.01 N 4.49. Found: C 63.15 H 5.12 N 4.23.

5,10,15,20-Tetrakis(p-α/β-D-galactopyran-6'-oloxycarbonylphenyl)porphyrin (4e)

According to the procedure given above 35 mg (0.018 mmol) 3e were reacted. The reaction mixture was poured into water (40 ml) and neutralised with 20% Na₂CO₃ solution. The porphyrin was isolated by filtration (0.45 μm microfilter) washed with water and ethanol and dried. Yield 26 mg (91%) 4e.- ¹H NMR (DMSO-d₆): δ = -2.95 (br s, 2 H, NH), 3.76 - 3.95 (m, 16 H), 4.32 - 4.58 (m, 16 H), 4.71 - 4.79 (m, 4 H), 5.02 (m, 2 H, 1'-H_o), 5.25 (m, 2 H), 6.29 (br m, 2 H, OH), 6.65 (br m, 2 H, OH), 8.40 (m, 16 H, H_o, H_m), 8.85 (br s, 8 H, H_β).- ¹³C NMR (DMSO-d₆): δ = 65.04 (t, C-6'), 65.23 (t, C-6'), 66.84 (t, C-6'), 67.91 (d), 68.14 (d), 68.91 (d), 69.01 (d), 69.38 (d), 69.64 (d), 72.23 (d), 72.31 (d), 73.45 (d), 74.89 (d), 76.39 (d), 77.72 (d), 81.77 (d), 82.11 (d), 83.04 (d), 93.11 (d, αC-1'), 95.85 (d), 97.32 (d), 97.86 (d, βC-1'), 102.18 (d, C_m), 119.51 (s, C-5, C-10, C-15, C-20), 128.16 (d, C_m), 129.71 (s, C_p), 132.00 (d, C_β), 134.82 (d, C_o), 146.15 (s, C_i), 147.69 (br s, C_α), 166.07 (s, CO).- IR (KBr): ν = 3384 (OH), 1707 (CO) cm⁻¹.- UV-Vis (H₂O): λ = 418, 516, 554, 591, 645 nm.- FAB-MS: m/e = 1438 (M⁺)- Anal. Calc. for C₇₂H₇₀N₄O₂₈ (1439.4): C 60.08 H 4.90 N 3.89. Found: C 59.66 H 4.75 N 3.88.

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10. Besides the α - and β -isomers, the NMR spectra indicate the existence of several conformers of compounds **4a** - **4e**. The existence of these isomers is due to rotation around the C-6'-O bond and rotation around the CO-O bond.

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